ADAR2-mediated editing of RNA substrates in the nucleolus is inhibited by C/D small nucleolar RNAs

Patrice Vitali, Eugenia Basyuk, Elodie Le Meur, Edouard Bertrand, Françoise Muscatelli, Jérôme Cavaillé, and Alexander Huttenhofer

1Laboratoire de Biologie Moléculaire des Eucaryotes, Centre National de la Recherche Scientifique (CNRS), UMR 5095, Institut Fédératif de Recherche (IFR) 109, 31062 Cedex Toulouse, France
2Institut de Genetique Moleculaire de Montpellier, CNRS, UMR 5535, Université Montpellier II, IFR 122, 34293 Montpellier Cedex 5, France
3Innbruck Biocentre, Division of Genomics and RNomics, Innsbruck Medical University, 6020 Innsbruck, Austria
4Institut de Biologie du Développement de Marseille, Campus de Luminy, 13288 Cedex 9, Marseille, France

Introduction

Primary RNA transcripts can undergo adenosine to inosine (A-to-I) RNA editing through hydrolytic deamination at the C-6 atom of adenosine (for reviews see Bass, 2002; Schaub and Keller, 2002; and Maas et al., 2003). These posttranscriptional RNA processing events are catalyzed by specific enzymes called adenosine deaminases that act on RNA (ADARs; Keller et al., 1999; Bass, 2002). Three structurally related members of the ADAR family have been characterized: ADAR1 and 2, which are ubiquitously expressed in many tissues with strong expression in the brain (Wagner et al., 1990); and ADAR3/RED2, which is only expressed in neuronal tissues and has not been proven to be an active enzyme (Melcher et al., 1996a; Chen et al., 2000). A-to-I RNA editing occurs within completely or partially double-stranded (ds) RNA structures. Sites of A-to-I editing are generally found within non-coding sequences such as repetitive sequences, introns, and 3’ untranslated regions (Morse et al., 2002; Levanon et al., 2004). However, a few site-specific edited sites have been also detected within the open reading frame of cellular and viral pre-mRNA transcripts. Because the newly introduced inosine residue is decoded as a guanosine by the translation apparatus, A-to-I RNA editing has the potential to diversify the genetic information so that more than one protein is produced from a single gene (Bass, 2002; Schaab and Keller, 2002; Maas et al., 2003; Seeburg and Hartner, 2003).

In the best understood examples of site-specific A-to-I RNA editing in mammals, ionotropic glutamate receptors (GluRs) and serotonin receptors (5-HT2C isoforms), the functional properties of the respective gene products, are modulated (Sommier et al., 1991; Burns et al., 1997; Seeburg and Hartner, 2003). In the case of GluR, RNA editing at the Q/R site of the GluR-B subunit of an α-amino-3-hydroxy-5-methyl-4-isoxazolepropionate receptor (a Gln codon is changed to an Arg codon) decreases the Ca2+ permeability of the receptor (Sommier et al., 1991; Burns et al., 1997; Seeburg and Hartner, 2003). In the case of GluR-B RNA editing at the Q/R site of the GluR-B subunit of an α-amino-3-hydroxy-5-methyl-4-isoxazolepropionate receptor (a Gln codon is changed to an Arg codon) decreases the Ca2+ permeability of the receptor (Sommier et al., 1991; Seeburg, 1996). Another edited site within GluR-B RNA is the R/G site (an Arg codon is changed to a Gty codon), resulting in an increase in the rate at which the channels recover from desensitization (Lomeli et al., 1994). In the serotonin 5-HT2C receptor pre-mRNA, five closely spaced ed-
ited adenosines (A–E sites) alter the coding potential of three codons (see Figs. 1 A and 3 A). By decreasing the efficiency of G-protein coupling, the RNA editing of 5-HT2C gives rise to serotonin receptor isoforms with a reduced constitutive activity (Burns et al., 1997; Herrick-Davis et al., 1999; Niswender et al., 1999; Berg et al., 2001).

The mechanism and the spatio-temporal regulation of site-specific RNA editing (Lomeli et al., 1994; Burns et al., 1997; Barbon et al., 2003; Yang et al., 2004) is still poorly understood. Most site-specific RNA editing requires a partial dsRNA structure that involves the exonic sequence encompassing the edited site(s) and a downstream intronic sequence (Higuchi et al., 1993; Burns et al., 1997). Thus, RNA editing and RNA splicing might interfere with each other or might even be coordinated (Higuchi et al., 2000; Wang et al., 2000; Bratt and Ohman, 2003; Flomen et al., 2004). Accordingly, ADAR1 and 2 are found to be associated with the spliceosome in a large nuclear particle, which is consistent with a physical interaction between editing and splicing machineries (Raitskin et al., 2001). Cellular annealing and unwinding activities affecting the overall stability of the dsRNA structure, which is recognized by ADARs, also play key roles to control RNA splicing and/or RNA editing (Reenan et al., 2000; Bratt and Ohman, 2003). Regulation of the intracellular levels of ADARs is expected to affect the overall efficiency of RNA editing (Kawahara et al., 2003; Yang et al., 2003a). In this line, ADAR2 is able to edit its own pre-mRNA to create a new 3′ splice site, the utilization of which results in a frameshift that gives rise to a nonfunctional ADAR2 protein. This autoregulative feedback loop might regulate the endogenous level of ADAR2 (Rueter et al., 1999). Surprisingly, the treatment of mice with fluoxetine (a serotonin re-uptake inhibitor) affects the RNA editing of 5-HT2C pre-mRNA, suggesting that RNA editing can be regulated in a serotonin-dependent manner (Gurevich et al., 2002).

In vivo, the subnuclear location where A-to-I RNA editing occurs has not been precisely characterized yet. However, because RNA editing reactions require the formation of an RNA duplex formed by base pairing between intronic and exonic sequences, editing has to occur before RNA splicing. Surprisingly, two observations suggest that nucleolar-associated functions might regulate RNA editing to some extent. First, ADAR1 and 2 are dynamically associated with the nucleolus, and, although a functional role of the nucleolus in the editing process is far from being understood, it has been proposed that nucleolar sequestration of the editing enzymes modulates RNA editing (Desterro et al., 2003; Sansam et al., 2003; Yang et al., 2003b; Nie et al., 2004). Also, we have previously identified a brain-specific C/D RNA, designated as MBII-52, that is predicted to target an A-to-I editing site within the 5-HT2C serotonin receptor mRNA for 2′-O-methylation of the ribose of the edited base. In mammals, site-specific 2′-O-methylations of RNAs (ribosomal RNAs) and U spliceosomal RNAs are directed by C/D RNAs, with most of them accumulating and functioning in the nucleolus (hence their designation as small nucleolar RNAs [snoRNAs]) and a few of them in the Cajal bodies (CBs; hence their designation as small CB-specific RNAs [scaRNAs]; Kiss, 2002). These C/D RNAs act through the formation of a canonical RNA duplex in which the nucleotide to be 2′-O-methylated in the target RNA is paired to the fifth position upstream from the conserved D (or D′) box sequence motif found in all of the C/D small RNA (Cavaillé et al., 1996; Kiss-Laszlo et al., 1996). Importantly, the nucleotide that is predicted to be modified by MBII-52 C/D RNA is precisely the edited C-site within 5-HT2C pre-mRNA. A 2′-O-methylation of the ribose of an adenosine nucleotide has been shown to specifically decrease the deamination rate of the modified adenosine in vitro (Yi-Bruzzozi et al., 1999); thus, by its antisense element, MBII-52 is predicted to negatively regulate pre-mRNA editing at the C-site (Cavaillé et al., 2000).

In this study, we designed several experiments to address unresolved questions regarding the potential involvement of the nucleolar compartment in the control of site-specific RNA editing. Thereby, we wanted to specifically address the following points: (1) Are editing enzymes ADAR1 and 2 functional within the nucleolus? (2) Is MBII-52 a bona fide C/D methylation snoRNA guide? And (3) does expression of MBII-52 snoRNA affect RNA editing at the C-site of a 5-HT2C RNA substrate?

**Results**

**ADAR2, but not ADAR1, can edit nucleolar 5-HT2C and GluR-B dsRNA substrates**

Several groups have recently reported that endogenous ADAR1 and 2 display a dynamic nucleolar association, with ADAR2 shuttling between the nucleolus and the nucleoplasm and ADAR1 shuttling between the nucleolus, the nucleoplasm, and the cytoplasm (Desterro et al., 2003; Sansam et al., 2003; Yang et al., 2003b; Nie et al., 2004). Although the functional significance of these observations is unknown, it has been proposed that RNA editing might be regulated by the intracellular compartmentalization of these ADARs (Desterro et al., 2003; Sansam et al., 2003). To investigate whether ADARs are active within the nucleolus, we analyzed RNA editing within a 5-HT2C dsRNA substrate (A–E-sites) and two GluR-B dsRNA substrates (Q/R and R/G sites), transcribed either from a cytomegalovirus (CMV) promoter (Pol II) or a ribosomal gene promoter (Pol I; Fig. 1, A and B). We reasoned that the distinct subcellular location expected for Pol II and Pol I transcripts (i.e., in the nucleoplasm and the nucleolus, respectively; Fig. 1 C) allows discrimination between nucleolar and nucleoplasmic RNA editing. Indeed, previous studies have shown that the majority of Pol I transcripts do not leave the nucleolus to a large extent (Cavaillé et al., 1996; Ganot et al., 1999; Jady et al., 2003). Pol I- and Pol II-5-HT2C minigenes were, therefore, transiently transfected into NIH-3T3 (which contains low levels of endogenous editing activities) together with either hADAR1- or hADAR2-expressing vectors. RNA editing efficiency was measured at each edited site by direct sequencing of the RT-PCR products (Fig. 2 A). Consistent with earlier reports (Burns et al., 1997; Higuchi et al., 2000; Hartner et al., 2004), when nucleoplasmic RNA editing was assayed, we observed that RNA editing at the D-site is mainly...
catalyzed by ADAR2, whereas ADAR1 preferentially edits the A- and B-sites; E- and C-sites are catalyzed by both enzymes (Fig. 2 A). In the course of this work, we also detected a novel edited site: the G-site edited by ADAR2 (Fig. 2 A), which maps within an intronic segment that base pairs with the exonic sequence containing the A–E-sites (Fig. 1 A). Intriguingly, this novel edited site was not detected in total RNA that was extracted from mouse brains (unpublished data); thus, its biological relevance is questionable.

Surprisingly, when nucleolar RNA editing was assayed, only adenosines at the D- and G-sites (edited by ADAR2) were found to be significantly converted into inosines (58.01 ± 4.6% and 15.3 ± 3.2%, respectively), whereas A- and B-sites (edited by ADAR1) remained fully unedited. Nevertheless, sequencing of the cloned RT-PCR products obtained from the same transfection experiments indicated that ~5% of the C-site was edited by ADAR2 in the nucleolus (unpublished data). Therefore, we reanalyzed RNA editing at the C-site by the more sensitive primer extension method (Burns et al., 1997). As shown in Fig. 2 A (right), nucleolar RNA editing mediated by ADAR2 is readily detected at the C-site (3.9 ± 0.63%) and also at the E-site, albeit at very low levels.

We also tested whether nucleolar RNA editing can occur at the Q/R and R/G edited sites in GluR-B dsRNA substrates. Again, we found that the Q/R site (only edited by ADAR2 [Higuchi et al., 2000]) as well as the R/G site (edited by ADAR1 and 2 [Maas et al., 1996; Melcher et al., 1996a,b]) are efficiently deaminated by ADAR2 in either Pol II or Pol I/GluR-B transcripts (efficiency of editing at Q/R and R/G sites in the Pol I transcripts is 40.1 ± 2.65% and 49.85 ± 13%, respectively; Fig. 2 B). In contrast, ADAR1 faithfully edits the R/G site in the Pol II/GluR-B dsRNAs, but not within Pol I transcripts, because it is targeted to the nucleolus (Fig. 2 B). Thus, we conclude that ADAR2 is active within the nucleolar compartment, whereas ADAR1 is not.

Given that Desterro et al. (2003) recently reported that an NH2-terminal deletion mutant of ADAR1 (lacking aa 1–442), termed GFP-ADAR1C-Term, is exclusively targeted to the nucleolus, we tested its ability to be active in RNA editing. Again, although the truncated GFP-ADAR1C-Term protein can edit the A- and B-edited sites as well as the R/G site in a Pol II–transcribed dsRNA substrate, no RNA editing could be detected within the same Pol I–transcribed dsRNA (Fig. 2 C). As controls, we monitored the distribution of the transfected GFP-tagged versions of ADAR2 and ADAR1C-Term. In agreement with an earlier report (Desterro et al., 2003), the two GFP-tagged proteins were mainly detected within the nucleolus (Fig. 2 D). Altogether, these data indicate that although both GFP-ADAR1C-Term and ADAR2-GFP proteins are detected in the nucleolus, only ADAR2 is able to efficiently edit a dsRNA structure within this organelle. It is noteworthy that endogenous levels of ADAR2 in NIH-3T3 do not support an efficient nucleolar RNA editing (Fig. 2 A, right). This is in contrast to the neuroblastoma cell line Neuro2A (unpublished data), suggesting that the intracellular level of endogenous ADAR2 protein might be critical for the nucleolar activity of the enzyme.

**MBII-52 exhibits all of the functional properties of a C/D nucleolar methylation RNA guide**

In addition to a potential role of the nucleolus in sequestering ADAR1 and 2 activities (Desterro et al., 2003; Sansam et al., 2003) and/or representing one of the nuclear sites of ADAR2-mediated RNA editing (Fig. 2), nucleolar-associated functions might also be required to fine tune RNA editing in a sequence-specific manner. Consistent with that assumption, MBII-52 (a C/D small RNA) displays a long antisense element matching a nucleotide tract within 5-HT2C pre-mRNA. This undergoes both alternative splicing (Canton et al., 1996; Wang et al., 2000) and A-to-I RNA editing (Burns et al., 1997), thus raising the possibility that this small RNA might control 5-HT2C expression (Cavaillé et al., 2000; Fig. 3 A). To get better insight into that hypothesis, we first asked whether this C/D RNA belongs to the snoRNA that accumulates within the nucleolus or...
to the scaRNA, a recently identified class of C/D RNAs that are only detected within the CBs (Kiss, 2002). In situ hybridizations with specific Cy3-labeled fluorescent oligonucleotide probes against MBII-52 was performed on adult rat brain sections. As shown in Fig. 3 B (left), MBII-52–specific signals merge perfectly with those of U3 C/D snoRNA, which is known to reside in the nucleolus. In addition, in many cells, this probe also gives rise to a more punctate nucleoplasmic signal outside the nucleolus (Fig. 3 B, right) that has been identified as CBs based on colocalization with coilin, a marker of this nuclear body. Note that MBII-85 is present in the nucleolus of the cell as well (Fig. 3 B). Immunoprecipitation performed from a rat brain total extract also reveals that these two C/D RNAs interact with three common C/D RNA-binding proteins: namely, fibrillarin, NHPX, and NOP58 (Fig. 3 C).

We next examined whether MBII-52 can act as an RNA methylation guide by expressing a predicted, minimal RNA target (Fig. 3 A). A short fragment that was complementary to the antisense elements of MBII-52 was placed under the transcriptional control of a Pol I promoter (Pol I-52 target), which allows specific targeting of the RNA substrate to the nucleolus (Ca-vaillé et al., 1996; Fig. 1). The plasmid construct was transiently transfected into NIH-3T3 cells in the presence or absence of an MBII-52–expressing vector. The presence of the targeted 2′-O-methylation was then assayed by primer extension analysis, as described previously (Maden, 2001). As shown in Fig. 3 D, a deoxynucleotide triphosphate (dNTP) concentration-dependent pause of reverse transcriptase was mapped at the expected nucleotide only in total RNA that was extracted from NIH-3T3 cotransfected with MBII-52–expressing plasmids. The same vector was also introduced into Neuro2A cells, a neuroblastoma cell line that endogenously expresses MBII-52, albeit at low levels (unpublished data); again, the same dNTP concentration-dependent pause pattern was observed. We conclude that the endogenously expressed MBII-52, as well as MBII-52 expressed from a transfected plasmid, can direct a sequence-specific 2′-O-methylation into an RNA target. Hence, MBII-52 can be considered as a bona fide methylation guide C/D snoRNA.

**Partial coexpression of MBII-52 and 5-HT2C serotonin receptor mRNA in the adult mouse brain**

A prerequisite for the in vivo role of MBII-52 in the editing of 5-HT2C receptor pre-mRNA is to show the coexpression of
both transcripts in the same brain regions. Thus, we performed in situ hybridization on adjacent sections of adult mouse brains with probes specific for MBII-52 and 5-HT2C receptor pre-mRNA to compare their expression patterns. To reveal trace levels of serotonin 5-HT2C expression (which could not have been detected with a 5-HT2C receptor antisense riboprobe), we also used a probe for the H/ACA snoRNA MBI-36. This small RNA is specifically encoded within intron 2 of 5-HT2C pre-mRNA (Fig. 3 A) and accumulates as a stable product concentrated in the nucleus; thus, it was easily detected. MBII-52 is expressed only in neurons of a broad variety of adult mouse brain structures—including the dentate gyrus; CA1, CA2, and CA3 in the hippocampus, granular cell layer, mitral cell layer, and glomerular cell layer in the olfactory bulb; or Purkinje cells in the cerebellum (unpublished data)—but is never expressed in the choroids plexus, whereas the 5-HT2C receptor and MBI-36 are expressed within this structure (Fig. 4, A–C). This is consistent with earlier reports (Molineaux et al., 1989; Cavaillé et al., 2000). However, among some structures where MBII-52 is expressed, we also detected an expression of 5-HT2C and MBI-36 (Fig. 4, D–L). Indeed, in the subthalamic nucleus (Fig. 4, D–F) and the piriform cortex (Fig. 4, G–I), MBII-52, 5-HT2C receptor, and MBII-36 display relevant, overlapping expression patterns. In some other brain structures, such as the medial and lateral parts of the geniculate nuclei, MBII-52, 5-HT2C receptor, and MBII-36, expression patterns partially overlap with a more restricted expression of 5-HT2C receptor (Fig. 4, J–L). These results are consistent with the ability of MBII-52 to play a role in the processing of 5-HT2C pre-mRNA in the neurons of some adult brain structures.

Nucleolar, but not nucleoplasmic, RNA editing can be specifically inhibited by a targeted C/D snoRNA

Although it is an attractive model, the in vivo involvement of MBII-52 in the posttranscriptional regulation of 5-HT2C pre-mRNA expression is difficult to formally prove because: (1) no classical methodology is sensitive enough to reliably identify 2′-O-methylated nucleotides in low-abundance RNA; (2) the transient nature of the in vivo interaction between C/D RNA and its RNA target hampers the use of classical biochemical approaches; and (3) no easily tractable cell lines express MBII-52 and 5-HT2C genes at levels comparable with those found in brain tissues (unpublished data). We next asked whether the expression of C/D snoRNAs bearing an antisense element that is designed to target an adenosine to be edited can interfere with its deamination level, as proposed by the model that MBII-52 snoRNA targets 5-HT2C pre-mRNA. To test this possibility,
Discussion

In recent years, it has become evident that gene expression in eukaryotes is linked to spatial nuclear compartmentalization (Carmo-Fonseca, 2002; Chubb and Bickmore, 2003). One of the best known examples is the nucleolus, which plays a key role in ribosome biogenesis as well as in a broad range of cellular processes, RNA-processing steps, and nuclear functions (Olson et al., 2002). The nucleolus is a nonmembrane-bound nuclear structure, and although its organization is not completely understood, recent studies lead to the view that there is a constant flux of molecules between the nucleolus and its surrounding nucleoplasmic environment (Phair and Misteli, 2000; Andersen et al., 2005). In this line, ADARs, the enzymes that catalyze A-to-I RNA editing, display a complex subcellular compartmentalization, with ADAR2 being localized in the nucleolus as well as in the nucleoplasm, whereas ADAR1 locates throughout the nucleolus, nucleoplasm, and cytoplasm (Desterro et al., 2003; Sansam et al., 2003; Yang et al., 2003b; Nie et al., 2004). Photobleaching experiments indicate that ADARs are constantly and rapidly shuttling in and out of the nucleolus (Desterro et al., 2003; Sansam et al., 2003). Furthermore, upon the transfection of a Pol II transcript–containing edited site, both ADAR1 and 2 are excluded from the nucleoli, and this translocation step is correlated with an increased RNA editing in the nucleoplasm (Desterro et al., 2003; Sansam et al., 2003).

Overall, these data show that the nucleolar association of ADAR enzymes is transient, and the nucleolar environment might merely represent a storage compartment rather than a functional site of RNA editing. However, the RNA editing activities of nucleolar-localized ADAR enzymes have not been determined until now. Through a specific targeting strategy of dsRNA substrates to the nucleolus (Figs. 1 and 2), we showed that ADAR2- but not ADAR1-mediated RNA editing can occur in the nucleolus, raising the possibility that endogenous cellular and/or viral transcripts might be edited within this subnuclear compartment.

The inability of ADAR1 to perform nucleolar RNA editing is unexpected and deserves several comments. Because human ADAR1 and 2 localize in the same subnuclear compartment (Desterro et al., 2003), a subtle differential intranucleolar distribution between these two enzymes is unlikely to account for differences in their nucleolar editing activities. ADAR1 exhibits unique features compared with other ADAR enzymes, including two Z-DNA–binding protein domains and an additional (third) dsRNA-binding domain. Thus, ADAR1-mediated RNA editing might require strict coupling with the nucleoplasmic environment and/or with the transcribing Pol II apparatus. In this line, ADAR1 in Xenopus laevis associates with transcriptionally active lampbrush chromosomes, suggesting that RNA editing by ADAR1 might occur cotranscriptionally (Doyle and Jantsch, 2003). We also cannot rule out the possibility that ADAR1 is posttranscriptionally modified in the nucleolus, thus rendering it inactive (J. Desterro, personal communication). Finally, the nucleolar form of ADAR1 might be linked to other functions that are not directly related to RNA editing.
Another objective of this study was to investigate the functional relevance of the putative MBII-52/5-HT2C interaction, specifically its capacity to interfere with RNA editing. By localization and associated protein-binding partners, MBII-52 is a bona fide methylation guide C/D snoRNA (Fig. 3) that displays a partially overlapping expression pattern with its putative RNA target in some regions of the adult mouse brain (Fig. 4). Remarkably, our study shows that MBII-52 RNA constructs and artificial C/D snoRNAs reduce ADAR2-mediated nucleolar editing of an RNA substrate in a sequence-specific manner (Fig. 5), whereas nucleoplasmic RNA editing remains unaffected. Artificial C/D snoRNAs have been previously reported to target 2'-O-methylations at ectopic positions in rRNAs or in artificial RNAs (Cavaillé et al., 1996; Kiss-Laszlo et al., 1996; Liu and Fournier, 2004), but to our knowledge our study represents the first demonstration that expression of a cognate C/D snoRNA inhibits a specific RNA processing event in mammalian cells. Together, our results are consistent with a model in which MBII-52 has the potential to inhibit RNA editing in vivo.

We are making the assumption that endogenously expressed 5-HT2C pre-mRNA might be targeted by MBII-52, resulting in decreased RNA editing at the cognate C-site of pre-mRNA. This interaction might eventually fine tune the G-protein coupling of the encoded serotonin 5-HT2C receptor. Indeed, the edited C-site plays (together with the E-site) a pivotal role in regulating serotonergic signal transduction (Burns et al., 1997; Niswender et al., 1999; Wang et al., 2000; Gurvich et al., 2002). Our model implies an at least transient localization of 5-HT2C pre-mRNA in the nucleolus. How this would be achieved is not addressed in this study. It is noteworthy, however, that pre-mRNA splicing can sometimes be uncoupled from cotranscriptional events and can occur away from the transcription sites (Lopez and Seraphin, 2000). In addition, 5-HT2C pre-mRNA hosts an H/ACA snoRNA within intron 2 (Fig. 3 A), which could promote the targeting of intron-containing pre-mRNA to the nucleolus (a mechanism previously observed in plants; Shaw et al., 1998). In vivo interaction between MBII-52 and its putative RNA target is likely to be complex and could involve distinct cellular regulatory pathways, according to the brain regions in which both transcripts are expressed (Fig. 4). Thus, at this point, we cannot rule out the alternative possibility (e.g., that under some circumstances not mimicked by our experimental transfection assay) that MBII-52 C/D small nucleolar ribonucleoprotein particles might have the ability to function outside of the nucleolus.

The human homologues of MBII-52 genes are encoded at the imprinted 15q11-q13 chromosomal region. This locus is associated with the Prader-Willi syndrome (PWS), which is a neurobehavioral disorder characterized by a complex phenotype, including hyperphagia and behavioral problems with obsessive compulsive disorder (Nicholls and Knepper, 2001). Although the lack of expression of HBII-52 genes is not sufficient to result in PWS (Runte et al., 2005), it is noteworthy that a knockout mice model for 5-HT2C genes leads to epilepsy, to eating disorders as a result of abnormal control over feeding behavior (Tecott et al., 1995), and to compulsive behavior symptoms (Chou-Green et al., 2003) that are described in PWS patients. Hence, it is tempting to speculate that defects of HBII-52 expression, which are presumably associated with the lack of expression of additional paternally expressed genes, might contribute to some aspects of the PWS disease.
Materials and methods

Unless otherwise noted, all techniques for cloning and manipulating nucleic acids were performed according to standard protocols.

Oligonucleotides

Modified and unmodified oligonucleotides were synthesized by Y. de Redon (Laboratoire de Biologie Moléculaire des Eucaryotes, Toulouse, France) on a PerSeptive Biosystems Expedite apparatus or at the MWG Biotech AG.

Plasmid constructs

5-HT2C and Glur8 dsRNA substrates were PCR amplified from rat and mouse genomic DNA, respectively, and were cloned into pBluescript KS+ (Stratagene) between the EcoRI and KpnI restriction sites or into pBluescript II SK+ (Stratagene) between the EcoRI and HindIII restriction sites. The same PCR products were also inserted under the control of a CMV promoter into pcDNA3.1 (Invitrogen) to generate Pol I/S-HT2C, Pol II/Glur8 (Q/R), and Pol II/Glur8 (R/G) vectors. The same PCR products were also inserted under the control of a CMV promoter into pcDNA3.1 (Invitrogen) to generate Pol II/S-HT2C, Pol II/Glur8 (Q/R), and Pol II/Glur8 (R/G). Minigene Pol I-S2 target was derived from pBluescript II SK+ (Stratagene) between the EcoRI and HindIII restriction sites, which were underlined sequence pCMV snRNA αoxy, pCMV snRNA αoxy/Q/R, and pCMV snRNA αoxy/R/G were all derived from pCMV-MBII-52 vectors (Cavaillé et al., 2000). The mammalian expression vector constructs of hADAR1 (pCS2-FLIS6-hADAR1) and hADAR2 (pCS2-FLIS6-hADAR2a) were gifts from W. Préval (Laboratoire de Biologie Moléculaire des Eucaryotes, Toulouse, France). The mammalian expression vector constructs of hADAR1 (pCS2-FLIS6-hADAR1) and hADAR2 (pCS2-FLIS6-hADAR2a) were gifts from W. Préval (Institut d’Informatique, Lisbon, Portugal).

Cell cultures and transfections

The murine NIH-3T3 and Neuro2A cells were cultured as monolayers in DME (GIBCO BRL) supplemented with 10% FBS (GIBCO BRL) at 37°C and 10% CO2. DNA for transfection assays was purified using a plasmid maxiprep kit (Qiagen). Subconfluent cells (~80% of confluency) were transiently transfected (DAC-30 reagent; Eurogentec) with 500 ng dsRNA substrate expressing plasmids and 4 μg D and snRNA-expressing vectors or 4 μg of empty pcDNA 3.1 (+) for negative controls.

Detection of ribose-methylated nucleotides

2′-O-methyl methylation was analyzed by reverse transcription by using low dNTP concentrations as described in Maden (2001), except that the lowest concentration was 0.01 mM dNTPs.

Analysis of RNA editing

48 h after transfection, total RNA was extracted using the TRIZol reagent (Invitrogen) according to the manufacturer’s protocol. Direct sequencing of RT-PCR products was performed using the BigDye terminator cycle sequencing reaction kit (PE Corp.) and was analyzed on a sequencer (model ABI Prism 377; PerkinElmer) using the Lasergene sequence analysis program package. Editing efficiency was quantified by measuring the height of peaks A (no edited site) and G (edited site) in order to determine the G/A + G peak ratio. This ratio was then reported to 100 to determine the percentage of editing at each site. All transfections were independently repeated three times to evaluate the SD. Primer extension assay was also performed as described in Burns et al. (1997) in the presence of 0.2 mM dATP, dCTP, and dGTP and 0.8 mM ddGTP and Taq polymerase (n = 50 cycles). Reactions were analyzed by gel electrophoresis on a 15% polyacrylamide gel, and intensities of the bands were quantified by Phospholmager (model FLA-3000; Fujititan). RNA editing levels at the C8 sites were determined by calculating the ratio of band intensity for each site to the sum of the three band intensities.

In situ hybridization

FISH on hybridized cells or rat brain sections was performed as described in Verheggen et al. (2002) by using oligonucleotide-specific probes for Pol I/S-HT2C RNA, Pol II/S-HT2C RNA, snoRNA MBII-52 and snoRNA MBII-85 (labeled with Fluorolink Cy3; Amersham Biosciences) and the U3-specific modified oligonucleotide with Fluorolink Cy5 (Amersham Biosciences). Nuclear DNA was stained by DAPI. Colorimetric in situ hybridization was performed on adult brains of C57BL/6 mice purchased from Iffa Credo as described in Tiveron et al. (1996). 5-HT2C receptor riboprobe (225 bp) hybridizing to the coding sequence of 5-HT2C receptor mRNA (nt 1060–1284 from GenBank/EMBL/DDB) under a pUC18 plasmid, and an 88-bp-long fragment of MBII-36 and a 77-bp-long fragment of MBII-52 were cloned, respectively. Probes were labeled with digoxigenin and were revealed with an antidigoxigenin antibody conjugated with AP.

Immunoprecipitation, immunostaining, and antibodies

C/D small nucleolar ribonucleoprotein particle immunoprecipitations from rat brain extracts have been performed as described in Cavaillé et al., (2001) with monoclonal antibibrillarin 72B9 (a gift of M. Pollard, W.M. Keck Autoimmune Disease Center, La Jolla, CA), rabbit anti-NHPX R66 (a gift of L.B. Chen, Harvard University, Boston, MA), and anti-NOP58 (a gift of R. Uhrmann, Max-Planck-Institute of Biophysical Chemistry, Gottingen, Germany). Immunostaining on rat brain sections have been carried out as described in Cavaillé et al. (2001) with a polyclonal anticoilin antibody (diluted 1:100; a gift of A. Lamond, Wellcome Trust Biocentre, Dundee, UK).

References


